Trace Element Imbalances in Hair and Nails of Alzheimer's Disease Patients

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ABSTRACT: The concentrations of 17 elements in the hair and nails of 180 Alzheimer's disease (AD) and control subjects have been determined by instrumental neutron activation analysis (INAA). Comparisons of trace element levels of properly matched AD and control groups revealed significant imbalances in the concentrations of six elements (Br, Ca, Co, Hg, K, and Zn) between disease and control groups. It is noteworthy that each of these has previously been shown by our group, or others, to be altered in some AD brain region(s). Geometric means for each element in both hair and nails of AD and control subjects are presented, and significant differences noted. The significance of these alterations with regard to the possible role of trace elements in the etiology of AD is discussed.

Key Words: Trace Elements, Alzheimer's Disease, Hair, Nail, Radiochemical Analysis

INTRODUCTION

Despite intense research efforts, the cause of Alzheimer's disease (AD) remains unknown. One of the hypotheses regarding its etiology implicates trace elements. A recent publication (Ehmann et al., 1986) summarized the elements that have been implicated in AD, with pro and con literature citations. This same study demonstrated imbalances in eight elements in AD brain compared to age-matched controls. In light of these findings, a follow-up study was initiated to determine whether or not the trace element imbalances observed in brain occurred in other tissues. Hair and nail results are reported here.

Many extensive reviews of the advantages, disadvantages, uses, and misuses of hair and nail analysis have appeared in the literature (Hopps, 1977; Valkovic, 1977; Laker, 1982;

Taylor, 1986; Evans and Jervis, 1987). Hair and nail were chosen for this study because they have been shown to reflect environmental and dietary exposure to many of the elements we were interested in, such as Hg (Phelps et al., 1980; Inasmasu et al., 1986); As (Valentine et al., 1979); Cd (Huel et al., 1984); Pb (Creason et al., 1975); Sb (Chattopadyhay and Jervis, 1974); and Cr (Rabinowitz et al., 1983). There is also precedent for using hair to monitor trace element changes in disease states. Alterations have been seen in hair Ca levels of myocardial infarction patients (Bacso, 1984); in Cd, Pb, and Zn levels in hypertension (Medeiros and Pellum, 1984); in Ca, Cl, I, Na, Mn, Se, and Zn in cancer (Thimaya and Ganapathy, 1982; Moo and Pillay, 1983); in Al and Ca in chronic renal failure (Marumo et al., 1984); and in Ca of Downs syndrome patients (Barlow et al., 1981).

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Few comprehensive studies have been done on trace element levels in normal nail tissue (Kanabrocki et al., 1979; Biswas et al., 1984), or for alterations in disease states (Djaldetti et al., 1987a, 1987b, Van Noord et al., 1987). This work provides a considerable addition to existing nail trace element data.

The use of hair and nail as trace element indicators also has its pitfalls. Trace element analysis of a single hair or nail sample from one individual generally does not provide useful information. A more acceptable use of hair as a bio-indicator involves comparisons between well-matched groups of subjects (Evans and Jervis, 1987). When large numbers of samples are used, the effects of individual variations are minimized, and any differences that are observed between the matched groups can be more reasonably correlated with a single factor, such as disease state.

The analytical technique used for elemental determinations in this study was instrumental neutron activation analysis (INAA). This method, with its sensitive and selective detection of many elements in diverse matrices, is well suited for trace element determinations in biological tissue. Little sample pretreatment is needed, so contamination is minimized. Additional discussion of the advantages and disadvantages of INAA for biological trace element work can be found elsewhere (Parr, 1980; Katz, 1985).

MATERIALS AND METHODS

The determination of hair and nail trace element levels and the subsequent nterpretation of the data are not simple inalytical problems. In this work, careful ittention was given to minimizing sample contamination in all aspects of the analytical procedure. All implements which contacted he samples were pre-washed in an ultra-sonic leaner using J. T. Baker Instra-Analyzed nitric cid, followed by rinsing with deionized listilled water and Distilled-in-Glass methanol. he polyethylene vials and bags used for ample storage and washing are very low in race elements (Moody and Lindstrom, 1977). laminar flow hood was used for sample ackaging, and talc-free gloves were used for Il sample handling.

Trace element levels in hair and nail are

known to be subject to a number of external factors, such as age, sex, hair treatment, and geographic location. Information on all these factors, and on health status, diet, and medication, was obtained from each patient. A separate study examined the effects of age, sex, and treatment on trace element concentrations (Vance et al., 1988), and only carefully matched disease-control groups were used for this study. Hair samples that had undergone any treatments (e.g., dyes, permanents, or rinses) or had been washed with Se- or Zncontaining shampoos were excluded from the study, as were samples from persons who noted any unusual exposure to trace elements in their work or home environments.

Hair and nail samples were obtained from AD and control subjects through the Sanders Brown Center on Aging at the University of Kentucky. The clinical diagnosis of AD was made according to criteria of the NINCDS/ADRDA work group (McKhann et al., 1984). The population consisted of 117 control and 63 AD subjects. All AD subjects were over the age of 45; 38 were female and 25 were male. The average age of all the AD patients was 71 years. 73% of the control group was over the age of 45; of these, 55 were female and 30 were male. The average age of the over-45 control group was 68 years.

Hair was removed from the nape of the neck of the subject using stainless steel scissors and fingernails were removed using scissors or stainless steel fingernail clippers. Samples were placed into individual Ziploc bags for temporary storage. In preparation for analysis, the 5 cm of hair proximal to the scalp was cut into pieces of approximately 1 cm using stainless steel scissors, and placed in virgin polyethylene scintillation vials for washing.

Nail samples were individually scraped with a synthetic quartz knife, cut if needed, and also placed in scintillation vials. The samples were washed according to the International Atomic Energy Agency (IAEA) method (IAEA, 1978), which consists of successive acetone and water washes with mechanical shaking, and allowed to air dry in a dust-free hood. Distilled-in-Glass acetone (J.T. Baker) and deionized distilled water were used for this procedure. It is recognized that the IAEA wash procedure can alter the levels of the more labile elements, such as the alkali metal cations. Multivalent transition element cations are, however,

generally more tightly bound to proteins and are less likely to be altered by the IAEA wash procedure.

Samples and standards were packaged in prewashed Suprasil quartz vials and subjected to a 40-hour irradiation at the Missouri Universities Research Reactor (MURR). All vials were counted immediately upon return from MURR for 20 min. to determine As, Au, Br, K, and Na. They were counted again approximately 10 days later for Ag, Ca, Co, Cr, Cs, Fe, Hg, Rb, Sb, Sc, Se, and Zn. Ortec HPGe and Ge (Li) detectors were used for counting. Gammaray spectra were acquired and analyzed using an ND680 multichannel analyzer system. Appropriate corrections were made for deadtime differences and spectral interferences.

Primary comparator standards used in this work were National Bureau of Standards Standard Reference Materials 1566 (Oyster Tissue), 1571 (Orchard Leaves), and 1577 (Bovine Liver). The secondary standard was Bowen's Kale. The results obtained for Bowen's Kale agree very well with accepted values (Vance, 1986). Samples of our Bowen's Kale had been analyzed previously in our laboratory using standard solutions prepared from high purity elements or stoichiometric compounds (Nadkarni and Ehmann, 1969).

RESULTS

The data were analyzed using the SAS Lifereg procedure (SAS Institute, 1985). This program was used because it provides a method by which detection limit values can be meaningfully incorporated into summary statistics. A more detailed description of the procedure has been provided elsewhere (Vance et al., 1988). Geometric means are reported (except for Zn), since most trace element concentrations in hair and nail are log-normally distributed.

The results of the hair and nail analyses are presented in Tables 1 and 2, respectively. Important external factors that affected the trace element concentrations in hair and nail (age, sex, treatments) had been identified previously (Vance et al., 1988), and these factors were taken into account when comparing disease and control groups. For each element geometric means for properly matched AD and control

groups are given, and significant differences (p. < 0.05) between the groups noted. Table 3 presents an overview of the trace element. imbalances observed by this laboratory in AD brain, hair, and nail.

There were no significant differences between AD and control groups for nine elements: Ag, As, Au, Cr, Fe, Na, Sb, Se, and Sc. The elements Cs and Rb were actually determined in less than 5% of the samples and these elements are not included in the statistical analysis. With regard to Na and K, we recognized the problems associated with measurement of these water-soluble cations. Our assumption was that the strictly controlled wash procedure should affect all samples in the same way, and that comparison studies should still be valid.

Hair

Four elements (Br, Ca, Co and Zn) were found to be imbalanced in the hair of AD subjects as compared to matched controls.

Bromine was higher (p < 0.05) in the hair of AD patients than in controls. Only hair that had not been treated with dyes, permanents, or rinses was used for this comparison. There were no age or sex effects for Br in hair.

Calcium was lower (p < 0.05) in the hair of AD subjects than in matched controls. Comparisons were again made only for nontreated hair, since treatments result in greatly increased Ca levels. Comparison groups were also matched by age (those over and under the age of 45) and sex, since these factors also affect hair Ca levels.

Cobalt was lower (p < 0.01) in AD subjects than in controls. Here too, only nontreated hair was included, and groups were sex-

Zinc was higher (p < 0.05) in the hair of AD patients than in age-matched controls.

Nail

Four elements (Br, Hg, K, and Zn) were imbalanced in the nail of AD subjects. Two of these (Br and Zn) were also imbalanced in the same direction in AD hair. Potassium was rarely detected in the hair, so no nail/hair comparisons are possible. Mercury was not

TABLE 1. Disease/Control Comparisons in Hair.

Element,	Groups ¹	AD Mean	Number of Samples ⁴	Matched Control Mean	Number of Samples
Ag. ng/g	none	53.1 (1.23)	[32-12-0]	87.6 (1.15)	[72-17-1]
As, ng/g	none	26.0 (1.24)	[10-34-0]	10.3 (1.32)	[24-63-3]
Au, ng/g	м	15.3 (1.23)	[24-0-0]		
	F	23.1 (1.27)	[19-0-1]	13.2 (1.23) 35.3 (1.23)	[29-0-3] [54-0-4]
Br. μg/g ³	N	3.59 (1.12)	[39-0-0]	2.55 (1.11)	[68-0-1]
Са но/23	M. > 45, N F. > 45, N	83.5 (1.59) 148 (1.68)	[8-14-2] [7-5-3]	208 (1.23) 458 (1.34)	[15-9-0] [13-9-0]
Co, ng/g ²	M.N F.N	9.37 (1.29) 14.5 (1.35)	[17-7-0] [12-3-0]	19.2 (1.15) 30.4 (1.25)	[29-3-0] [35-2-0]
Cr, ng/g	none	195 (1.18)	[40-3-1]	212 (1.13)	[84-6-0]
Fe. μg/g	none	9.99 (1.15)	[31-13-0]	9.16 (1.10)	[77-12-1]
Hg, ng/g	M F	355 (1.19) 557 (1.25)	[22-1-1] [20-0-0]	439 (1.09) 525 (1.11)	[32-0-0] [58-0-0]
Na, μο/g	none	4.62 (1.16)	[42-2-0]	4.6 (1.10)	[87-1-2]
Sb, ng/g	N	25.7 (1.14)	[31-8-0]	30.0 (1,12)	[65-3-1]
Sc. ng/g	>45	0.50 (1.16)	[26-18-0]	0.39 (1.20)	[29-34-1]
Se. μg/g	> 45, N	0.631 (1.04)	[35-0-4]	0.550 (1.04)	[44-0-3]
žn, μg/g ³	>45	164 (5.7)	[41-0-3]	143 (6.3)	[61-0-3]

¹ These factors had been determined previously to have significant effects on the trace element concentrations.

Note: All means are geometric means (x/+ SEM), except for Zn which is an arithmetic mean (± SEM).

imbalanced in AD hair.

Bromine is elevated (p < 0.05) in the nail of AD subjects as compared to age-matched controls. Potassium (p < 0.01), and zinc (p < 0.05) are both higher in AD subjects than in controls. No age or sex effects were observed for the in action. for Hg in nail.

DISCUSSION

This study has demonstrated significant differences in trace element levels in hair and nail between AD and control groups, indicating that trace element imbalances in AD are not restricted to the brain. A major point of

N = only non-treated hair samples used for comparisons.

M = male. F = female.

> 45 = only subjects over age 45 used for comparisons.

 $^{^2}$ Significant difference between AD and control; p < 0.01

³ Significant difference between AD and control; p < 0.05

⁴ Number of actual-detection limit-missing values, respectively

interest is that all six of the elements exhibiting imbalances in hair and/or nail (Br, Ca, Co, Hg, K, Zn) have also been shown to be imbalanced in some AD brain regions, although the imbalances are not always in the same direction. The relationship of the peripheral trace element imbalances to those in

the brain, and to the disease process itself, is not known. Some or all of the observed imbalances could reflect a generalized systemic alteration intrinsic to the disease. Or, they may be non-specific changes resulting from the terminal disease state with its attendant malnutrition, immobilization, dehydration, and

TABLE 2. Disease/Control Comparisons in Nail.

Element,	Groups ¹	AD Mean	Number of Samples ⁴	Matched Control Mean	Number of Samples
Ag, ng/g	М	21.9 (1.32)	[7-18-0]	********	
	F	36.0 (1.32)	[18-19-1]	23.5 (1.29) 42.8 (1.26)	[17-24-1]
As, ng/g	>45	33.8 (1.17)	[14-48-1]	28.6 (1.18)	[26-59-0]
Au, ng/g	м	11.5 (1.31)	[25-0-0]		
	F	24.1 (1.12)	[34-2-2]	12.9 (1.30) 31.8 (1.15)	[36-2-4] [70-1-4]
Br, μg/g ²	>45	2.48 (1.87)	[63-0-1]	1.94 (1.05)	[83-0-2]
Са, µg/g	none	472 (1.08)	[35-16-12]	522 (1.06)	[70-32-15]
Co, ng/g	none	19.3 (1.15)	[51-11-1]	27.5 (1.10)	[96-20-1]
Cr. ng/g	none	1328 (1.13)	[62-0-1]	1760 (1.10)	[115-1-1]
•. нс/д	> 45	12.1 (1.11)	[46-15-2]	12.3 (1.10)	[66-18-1]
ta. na/g ³	none	132 (1.14)	[62-1-0]	170 (1.07)	[112-5-0]
, μg/g ² .	M, > 45	88.6 (1.27)			(
	F, >45	75.4 (1.22)	[14-9-2] [18-11-9]	50.0 (1.24) 25.9 (1.26)	[16-9-5] [22-24-9]
a. µg/g	M > 45	204 (1.16)			()
	F. > 45	125 (1.13)	[25-0-0] [38-0-0]	134 (1.12) 112 (1.08)	[30-0-0]
b, ng/g	>45	19.5 (1.13)	[40-22-1]	21.3 (1.11)	[60-25-0]
c, ng/g	none	1.10 (1.17)	[40-23-0]	0.76 (1.15)	
r, μg/g	м	0.004.44.4		()	[72-45-0]
	F	0.991 (1.07)	[25-0-0]	0.99 (1.05)	[41-0-1]
		1.081 (1.03)	[37-0-1]	1.10 (1.03)	[75-0-0]
. μg/g ³	M, >45	158 (5.9)			1 1
	F, >45	172 (5.3)	[25-0-0] [38-0-0]	145 (6.3) 161 (4.4)	[29-0-1] [54-0-1]

¹ These factors had been determined previously to have significant effects on the trace element concentrations,

N = only non-treated hair samples used for comparisons.

M = male. F = female.

> 45 = only subjects over age 45 used for comparisons.

² Significant difference between AD and control; p < 0.01

³ Significant difference between AD and control; p < 0.05

⁴ Number of actual-detection limit-missing values, respectively

Note: All means are geometric means (x/+ SEM), except for Zn which is an arithmetic mean (± SEM).

TABLE 3. Summary of Elemental Imbalances in Brain, Hair, and Nail of AD Patients.

	Brain		Nail	Hair
Element	Cortex ¹	Special Regions 2.3		
Br	elevated		elevated	
Ca	n.d.	n.d.	enevated	elevated
a		n.d.	n.d.	lowered
Со		elevated		n.d, lowered
Ca	lowered	(nbM) lowered	n. d.	n.d.
Hg	elevated	(h) elevated	lowered	
K	lowered	(nbM) lowered	elevated	n.d.
N -	(gray) lowered	(a,h)	nd	
Na	elevated	(a, nbM)		n.d.
Р	elevated	(all 3) lowered		
Rь		(a)	n.d.	n.d
	lowered	lowered (a,h)	n.d.	n.d.
Se	-	elevated (nbM)	*****	
Zn		elevated (a)	elevated	elevated

¹Ehmann et al., 1986.

so on. We offer here some speculations about the ways in which trace substances might be involved in the clinical, biochemical, and pathological alterations observed in AD.

Bromine

The elevation of Br in hair, nails, and brain of AD subjects has been a consistent imbalance in our studies. Akanle et al. (1987) did not observe a Br imbalance in their study of 9 senile dementia subjects and 17 controls, but they do not specify whether or not treated hair

samples were excluded from their study. Ward et al. (1987) found Br to be elevated in the hippocampus and cerebral cortex of AD subjects as compared to controls. Rindby et al. (1983) have reported an elevation of Br in the serum and CSF of four patients suffering from senile dementia.

Bromide has not been shown to have any essential physiological function, but it is known to be psychoactive, and was once used as a sedative and anticonvulsant. The amounts of Br to which humans are exposed has increased in this century due to increased use of

²Thompson et al., 1988.

³nbM = nucleus basais of Meynbert: h = hippocampus; a = amygdala. elevated = AD subjects have higher levels than matched controls (p < 0.05). lowered = AD subjects have lower levels than matched controls (p < 0.05).</p>

⁻ no difference between AD and control subjects.

n. d. = element not determined.

Note: Only imbalances observed in our laboratory are included.

Br-containing fumigants and increased mining wastes (Rauws, 1983). Though there are reports of bromide intoxication resulting from excess intake of bromide-containing medications (Trump and Hochberg, 1976; Kunze, 1976), exposure to environmental Br, even at higher than normal levels, seems to produce only minor neurobehavioral alterations (Anger et al., 1986).

Relatively little is known about the actual cellular mechanisms by which Br exerts its central nervous system effects. It can substitute for Cl or, to a lesser extent, for I, in cell processes. Sangster et al. (1983) have shown that interference of Br with, or substitution for, I can result in alterations in T₃ or T₄ levels. A link between thyroid disorders and AD has been reported (Heyman et al., 1984), although a relationship between Br, thyroid disorders and AD seems unlikely. Interference with Cl seems a more likely problem.

The replacement of CI by Br does seem to have an effect on neurotransmitter function. Roskoski (1974) showed that brominated components of the cholinergic system inhibit the action of CAT, an enzyme known to be affected in AD (Davies and Maloney, 1976). Goodwin et al. (1969) showed that the replacement of CI by Br in the perfusing medium of slices of rat striatum diminished the release of noradrenaline and serotonin. Serotonin has been shown to be decreased in AD (Sparks et al., 1986). Iodide did not produce this effect, suggesting that the inhibition was specific to Br. Because brain Cl levels are much higher than Br (Ehmann et al., 1986), it is questionable as to how much interference the Br could actually produce in brain, unless the Br were selectively utilized in place of Cl. This phenomenon has been shown to occur in eosinophils. Weiss et al. (1986) showed that human eosinophils would preferentially oxidize Br to a halogenating intermediate in the presence of at least 1000fold excess of Cl. HOBr is a powerful and toxic oxidant, reacting with a variety of biomolecules, and can act to halogenate proteins.

The Br elevation observed in hair and nail of AD subjects could also result from other causes. It has been shown that treatment with lithium carbonate results in elevated hair Br (Handorf et al., 1985; Campbell et al., 1986).

Perhaps other medications commonly taken by AD patients have similar effects. It has also been shown that Cl depletion greatly lengthens the elimination half-life of Br (Rauws, 1983). If AD subjects were on salt-restricted diets, the longer residence time of Br in the body could lead to increased Br levels in hair and nail.

Calcium

Hair Ca is decreased in AD patients compared to age-, sex-, and treatment-matched controls. The same trend toward lowered Ca was observed in nails, but was not significant. These results disagree with those of Shore et al. (1984) and Akanle et al. (1987) who reported no difference in hair Ca between AD and control subjects. However, in neither of these studies is the problem of the effects of hair treatment on hair Ca levels addressed. In a study of Downs' syndrome (DS) patients Barlow et al. (1981) reported that both male and female DS patients exhibited lowered hair Ca values compared to matched controls. It is known that DS patients develop dementia and brain morphological and biochemical changes similar to AD if they live to middle age (Wisniewski et al., 1985).

The lowering of hair Ca in AD subjects is opposite in direction to the elevation reported in AD brain (Hershey et al., 1985; Ward et al., 1987). This observation actually fits in with Gajdusek's proposal concerning the possible role of Ca in AD (Gajdusek, 1985). He proposed that a lowered amount of dietary and environmental Ca might result in malfunction of the parathyroid glands, with subsequent deposition of Ca in neurons. Gordus (1973) demonstrated that hair Ca levels reflect dietary intake. The lowered Ca intake could thus simultaneously result in lower hair Ca and elevated brain Ca.

Cobalt

This element is significantly decreased in the hair of AD subjects compared to controls. Nail Co levels are also lower in AD patients, but the difference is not significant.

There are few reports on Co levels in AD tissues. Cole and Prehal (1984) reported lowered serum Vitamin B₁₂ in AD subjects as compared to controls. Ehmann et al. (1986)

found no alterations in Co for most brain regions, but we did report that Co levels were elevated in the nucleus basalis of Meynert in AD (Thompson et al., 1988). Ward et al. (1987) reported no differences between AD and controls in Co levels in hippocampal and cerebral cortex samples.

Cobalt is not known to have neurotoxic effects. It does not seem plausible that Co has a direct role in AD.

Mercury

Mercury is decreased in the nail of AD subjects compared to controls. Though there are few studies in the literature on Hg in nail, many reports have indicated that hair is a good monitor for Hg exposure (Phelps et al., 1980; Inasmasu et al., 1986). Brain Hg levels have been reported to be elevated in AD by some workers (Ehmann et al., 1986; Thompson et al., 1988), but not by others (Ward et al., 1988).

Perhaps one reason for the lowering of nail Hg in AD subjects could be the presumably lower exposure rate of the AD patients to environmental Hg. This would not explain the elevated brain Hg, however. An alternative approach would be to consider that the disease process somehow alters the distribution of Hg in the body. For example, several researchers have speculated that the bloodbrain-barrier (BBB) is altered in AD (Wisniewski and Kollowski, 1982). It has been demonstrated that Hg ions will penetrate the BBB (Ware et al., 1974) and impair its normal function. If this occurred, then it might be possible that Hg could preferentially accumulate in brain and be depleted in other tissues that reflect recent environmental exposure.

Of the several elements seen to be imbalanced in AD brain, hair, or nail, Hg seems to have the most severe and well-documented neurotoxic effects on systems that are known to be affected in AD. One example of this is the documented decrease in protein synthesis and reduction of RNA levels in AD brain (Mann et al., 1982; Borthwick et al., 1985). These changes can be effected by the presence of Hg. Kim (1980) demonstrated decreases in protein content and in DNA and RNA levels in rats fed mercury salts. Sajdel-Sulkowska and Marotta (1984) used a mercury

compound to inactivate a protein that inhibits a cellular alkaline ribonuclease in brain. Decreased ribonuclease led to an increase in RNA degradation and a decrease in protein synthesis.

Mercury has also been demonstrated to have adverse effects on the function of Na*-K*-ATPase which in turn can lead to alterations in neurotransmitter uptake (Rajanna and Hobson, 1985). Hrdina et al. (1976) showed that exposure to methylmercury produced significant decreases in cortical ACh and brainstem 5-HT levels. Dwivedi et al. (1980) demonstrated a decrease in CAT activity in the brains of rats fed either methyl- or inorganic mercury.

Potassium

This alkali metal is elevated in the nails of AD patients as compared to their age- and sexmatched controls. Ehmann et al. (1986) reported a decrease in the separated gray/white matter of AD brain, and Ward et al. (1987) also noted a K decrease in AD hippocampus and cerebral cortex.

It is difficult to envision an alteration in K as a cause of AD. If anything, the change would be more likely to be a result of the disease process.

Zinc

This element is elevated in the hair and nail of AD subjects compared to controls. Ehmann et al. (1984) observed no Zn alterations in AD subjects in specimens derived principally from the cerebral cortex, but did find elevated Zn in the amygdala. Ward et al. (1987) reported decreased Zn levels in AD hippocampus and cerebral cortex.

Burnet (1981) suggested that the inability of neurons to incorporate Zn into DNA-handling enzymes could ultimately lead to dementia. This hypothesis does not necessarily imply that there is a Zn imbalance in bulk tissue, however.

It may well be that AD is not a simple disease that will eventually be traced to one cause, but rather is a complex disease with many interrelated causative factors. It is possible that an environmental factor such as a neurotoxic trace metal could serve as a trigger

in susceptible individuals and initiate a cascade of molecular changes resulting in the histopathological and biochemical alterations observed with AD.

In order to advance our understanding of the possible role of trace elements in AD, future studies will need to provide information on the localization of trace elements in the cell or pathological structure. An enhanced understanding of the biochemical interactions of non-essential elements like Br and Hg is also clearly needed.

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REFERENCES

- Akanle OA, Spyrou NM, Damyanov AA, Shaw DM, Ali L. Investigation of elemental models in senile dementia and depressives using neutron activation analysis. J Radioanal Nucl Chem 1987; 113:405-416
- Anger WK, Moody L, Burg J, Brightwell WS, Taylor BJ, Russo JM, Dickerson N, Setzer JV, Johnson BL, Hicks K. Neurobehavioral evaluation of soil and structural fumigators using methyl bromide and sulfuryl fluoride. Neurotoxicology 1986; 7:137-156
- Bacso, J. Short term and long term variations of calcium concentration in beard. J Radioanal Chem 1984; 83:167-173
- Barlow PJ, Sylvester PE, Dickerson JWT. Hair trace metal levels in Downs syndrome patients. J Ment Defic Res 1981; 25:161-168
- Biswas SK, Abdullah M, Akhter S,

- Tarafdor SA, Khaliquzzaman M, Khan AH. Trace elements in human fingernails: Measurement by proton induced x-ray emission. J Radioanal Nucl Chem 1984; 82:111-124
- Borthwick NM, Yates CM, Gordon A. Reduced proteins in temporal cortex in Alzheimer's disease: An electrophoretic study. J Neurochem 1985; 44:1436-1441
- Burnet FM. A possible role of zinc in the pathology of dementia. Lancet 1981; 1:186-188
- Campbell C, Ward NI, Peet M.
 Increased bromide levels in serum and hair
 during lithium treatment. J Affective
 Disord 1986: 11:161-164
- Chattopadyhay A, Jervis RE. Hair as an indicator of multielement exposure of populations groups. In: Trace Substances in Environmental Health, 8th Annual Conference, Hemphill, DD, ed. 1974, pp. 31-38
- Cole MG, Prehal JF. Low serum vitamin B₁₂ in Alzheimer-type dementia. Age and Aging 1984; 13:101-105
- Creason JP, Hinners TA, Bumgarner JE, Pinkerton C. Trace elements in hair as related to exposure in metropolitan New York. Clin Chem 1975; 21:603-612
- Davies P, Maloney AJR. Selective loss of central cholinergic neurons in Alzheimer's disease. Lancet 1976; 2:1403
- Djaldetti M, Fishman P, Harpaz D, Lurie B. X-ray microanalysis of the fingernails in cirrhotic patients. Dermatologica 1987a; 147:114-116
- Djaldetti M, Fishman P, Hart J. The iron content of fingernails in iron deficient patients. Clin Sci 1987b; 72:669-672
- Dwivedi C, Raghunathan R, Joshi BC, Foster HW. Effect of mercury compounds on cholineacetyltransferase. Res Commun Chem Pathol Pharmacol 1980; 30:381-384
- Ehmann WD, Markesbery WR, Alauddin M. Quantitation, localization and variations of brain zinc with aging by instrumental neutron activation analysis. In: Neurobiology of Zinc. Frederickson CJ, Howell GA, Kasarskis EJ, eds. New York, Alan R. Liss Inc., 1984; pp. 329-342
- Ehmann WD, Markesbery WR, Alauddin M, Hossain T, Brubaker

EH. Brain trace elements in Alzheimer's disease. Neurotoxicology 1986; 7:197-206

Evans GJ, Jervis RE. Hair as a bioindicator: Limitations and complications in the interpretation of results. J Radioanal Nucl Chem 1987; 110:613-625

Gajdusek DC. Hypothesis: Interference with axonal transport of neurofilaments as a common pathogenic mechanism in certain diseases of the central nervous system. New Eng J Med 1985; 312:714-719

Goodwin JS, Katz RI, Kopin IJ. Effect of bromide on evoked release of monoamines from brain slices and intact atria. Nature 1969; 221:556-557

Gordus A. Factors affecting the trace metal content of human hair. J Radioanal Chem 1973; 15:229-243

Handorf CR, Coleman JH, Rawls WN. Elevated serum bromide in patients taking lithium carbonate. J Clin Psychiat 1985; 46:9-10

Hershey CO. Hershey Wongmongkolrit T, Varnes AW, Breslau D. Trace element content of brain in Alzheimer's disease and aging. Trace Elem Med 1985; 2:40-43

Heyman A, Wilkinson WE, Stafford JA, Helms MJ, Sigmon AH, Weinberg T. Alzheimer's disease: A study of epidemiological aspects. Ann Neurol 1984; 15:335-341

Hopps HC. The biologic basis for using hair and nail for analyses of trace elements. Sci Total Environ 1977; 7:71-89

Hrdina PD, Peters DAV, Singhal RL. Effects of chronic exposure to cadmium, lead, and mercury on brain biogenic amines in the rat. Res Commun Chem Pathol Pharmacol 1976; 15:483-493

Huel G, Everson RB, Menger I. Increased hair cadmium in newborns of women occupationally exposed to heavy metals. Environ Res 1984; 35:115-121

Inasmasu T, Ogo A, Yanagawa M, Keshino M, Hrakoba A, Takahashi K, Ishinish N. Mercury concentration changes in human hair after the ingestion of canned tuna fish. Bull Environ Contam Toxicol 1986; 37:475-481

International Atomic Energy Agency. Activation Analysis of Hair as an Indicator of Contamination of Man by Environmental Trace Element Pollutants.

IAEA/RL/50. Vienna, Austria, IAEA, 1978

Kanabrocki EL, Kanabrocki JA, Greco J, Kaplan E, Oester YT. Instrumental analysis of trace elements in thumbnails of human subjects. Sci Total Environ 1979; 13:131-140

Katz SA. Determination of trace elements in biological tissues and fluids. American Biotechnology Laboratory 1985; 3:10-17

Kim MH. Effect of long term intake of mercuric chloride on memory behavior and contents of nucleic acid and catecholamines in rat brain. K'at'ollic Taehak Uihakpu Nonmumjip 1980; 33:279-91

Kunze U. Chronic bromide intoxication with a severe neurological deficit. J Neurol

1976; 213:149-152

Laker M. On determining trace element levels in man: The uses of blood and hair. Lancet 1982; 2:260-262

Mann DMA, Neary D, Yates PO, Lincoln J, Snowden JS, Stanworth P. Neurofibrillary pathology and protein synthetic capacity in nerve cells in Alzheimer's disease, Neuropathol Appl Neurobiol 1982; 8:161-176

Marumo F, Tsukamoto Y, Iwanami S. Kishimoto T, Yamagami S. Trace element concentrations in hair, fingernails, and plasma of patients with chronic renal failure on hemodialysis and hemofiltration.

Nephron 1984; 38:267-272

McKhann G, Drachman D, Folstein M, Katzman R, Price D, Stadlin EM. Clinical diagnosis of Alzheimer's disease: Report of the NINCDS-ADRDA work group. Neurology (NY) 1984; 34:939-944

Medeiros DM, Pellum LK. Elevation of cadmium, lead, and zinc in the hair of adult black female hypertensives. Bull Environ Contam Toxicol 1984; 32:525-532

Moo SP, Pillay KKS. Trace element profiles in the hair of cancer patients. J Radioanal Chem 1983; 77:141-147

JR, Lindstrom Selection and cleaning of plastic containers for storage of trace element samples. Anal Chem 1977; 49:2264-2267

Nadkarni RA, Ehmann Determination of trace elements in biological standard kale by neutron activation analysis. J Radioanal Chem

1969; 3:175-185

Parr RM (ed). Elemental Analysis of Biological Materials. Technical Report Series No. 197, IAEA, Vienna, 1980

Phelps RW, Clarkson TW, Kershaw TG, Wheatley B. Interrelationships of blood and hair mercury concentrations in a North American population exposed to methylmercury. Arch Environ Health 1980; 35:161-168

Rabinowitz MB, Gouick HC, Levin SR, Davidson MB. Clinical trial of chromium and yeast supplements on carbohydrate and lipid metabolism in diabetic men. Biol Trace Elem Res 1983; 5:449-466

Rajanna B, Hobson M. Influence of mercury on uptake of [3H]dopamine and [3H]norepinephrine by rat brain synaptosomes. Toxicol Lett 1985; 27:7-14

Rauws AG. Pharmacokinetics of bromide ion-an overview. Food Chem

Toxicol 1983; 21:379-382

Rindby A, Selin E, Standzenieks P. Application of an EDXRF-spectrometer in clinical investigations. In: Trace Elem Anal Chem Med Biol, Proc Int Workshop, 2nd, Berlin, Walter deGruyter and Co., 1983; 2:721-727

Roskoski R. Choline acetyltransferase: Reversible inhibition by bromoacetyl coenzyme A and bromoacetylcholine. Biochemistry 1974; 13:2295-2298

Sajdel-Sulkowska EM, Marotta CA. Alzheimer's disease brain: Alterations in RNA levels and in a ribonuclease-inhibitor complex. Science 1984; 225:947-949

Sangster B, Blom JL, Sekhuis VM, Loeber JG, Rauws AG, Koedam JC. The influence of sodium bromide in man: A study in human volunteers with special emphasis on the endocrine and the central nervous system. Food Chem Toxicol 1983; 21:409-419

SAS Institute, Inc. SAS User's Guide: Statistics, Version 5 Edition. Cary, North Carolina, SAS Institutes, Inc.

1985

Shore D, Henkin RI, Nelson NR, Agarwal RP, Wyatt RJ. Hair and serum copper, zinc, calcium and magnesium concentrations in Alzheimertype dementia. J Am Geriat Soc 1984; 32:892-895

Sparks DL, Markesbery WR, Slevin JT. Alzheimer's disease: Monoamine and spiperone binding reduced in nucleus basalis. Ann Neurol 1986; 19:602-604

Taylor A. Usefulness of measurements of trace elements in hair. Ann Clin Biochem

1986; 23:364-378

Thimaya S, Ganapathy Selenium in human hair in relation to age, diet, pathological condition and serum levels. Sci Total Environ 1982; 24:41-49

Thompson CM, Markesbery WR, Ehmann WD, Mao YX, Vance, DE. Regional brain trace-element studies in Alzheimer's disease. Neurotoxicology 1988; 9:1-8

Trump DL, Hochberg MC. Bromide intoxication. Johns Hopkins Med J 1976; 138:119-123

Valentine JL, Kang HK, Spivey G. Arsenic levels in human blood, urine, and hair in response to exposure via drinking water. Environ Res 1979; 20:24-32

Valkovic V. Trace Elements in Human Hair. Garland STPM Press, New York,

1977

Vance DE. Trace Element Relationships in Alzheimer's Disease: Hair and Nail Analysis by INAA. PhD Dissertation, University of Kentucky, 1986

Vance DE, Ehmann WD, Markesbery WR. Trace element content in fingernails and hair of a non-industrialized U.S. control population. Biol Trace Elem Res 1988, in press

VanNoord PAH, Collette HJA, Maas MJ, DeWaard F. Selenium levels in nails of premenopausal breast cancer patients assessed prediagnostically in a cohort-nested case-referent study among women screened in the DOM project. Int j Epidemiol 1987; 16:318-322

Ward NI, Mason JA. activation analysis for identifying elemental status in Alzheimer's disease. J Radioanal Nucl Chem 1987; 113:515-526

Ware RA, Chang LW, Burkholder PM. An ultrastructural study on the blood-brain barrier dysfunction following mercury intoxication. Acta Neuropathol (Berlin) 1974; 30:211-224

Weiss SJ, Test ST, Eckmann CM, Roos D, Regiani S. Brominating oxidants generated by human eosinophils. 208

Science 1986; 234:200-201
Wisniewski KE, Dalton AJ, Crapper-McLachlan DR, Wen GY,
Wisniewski HM. Alzheimer's disease
in Downs' syndrome: Clinicopathologic
studies. Neurology 1985; 35:957-961

Wisniewski HW, Kollowski PB. Evidence for blood-brain barrier changes in senile dementia of the Alzheimer type (SDAT). Ann NY Acad Sci 1982; 396:119-129